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MYOCARDIAL GRAFTS AND CELLULAR
COMPOSITIONS USEFUL FOR SAME

Instal *Instal* BACKGROUND OF THE INVENTION

The present invention resides generally in the field of cardiology, and more particularly relates to stable myocardial grafts and methods and cellular compositions useful for achieving such grafts.

As further background, organ transplantation has been widely used to replace diseased, nonfunctional tissue. More recently, cellular transplantation to augment deficiencies in host tissue function has emerged as a potential therapeutic paradigm. One example of this approach is the well publicized use of fetal tissue in individuals with Parkinsonism (reviewed in (1), see reference list, infra), where dopamine secretion from transplanted cells alleviates the deficiency in patients. In other studies, transplanted myoblasts from-uneffected siblings fused with endogenous myotubes in Duchenne's patients; importantly the grafted myotubes expressed wild-type dystrophin (2).

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Despite their relevance in other areas, these earlier studies do not describe any cellular transplantation technology which can be successfully applied to the heart, where the ability to replace damaged myocardium would have obvious clinical relevance. Additionally, the use of intra-cardiac grafts to target the long-term expression of angiogenic factors and ionotropic peptides would be of therapeutic value for individuals with myocardial ischemia or congestive heart failure, respectively.

In light of this background there is a need for the development of cellular transplantation technology in the heart. Desirably, such technology would not only provide stable grafts in the heart but also enable the delivery of useful recombinant proteins or other molecules directly to the heart. The present invention addresses these needs.

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SUMMARY OF THE INVENTION

The applicant has established cellular grafts in the myocardium which are viable long-term. Cardiomyocytes and skeletal myoblasts have been grafted directly into the myocardium of syngeneic animals. Viable grafts were detected at least one-half year post-implantation (the latest time point assayed). The presence of the grafts was not accompanied by overt cardiac arrhythmia, and the majority of the grafts were juxtaposed directly to the host myocardium and not encapsulated. It has thus been discovered that the myocardium can serve as a stable platform for cellular transplants. These transplants can be used for the local delivery of recombinant molecules to the heart and/or for replacing diseased tissue to supplement myocardial function.

Accordingly, one preferred embodiment of the invention provides a myocardial graft in an animal which includes a stable graft of skeletal myoblasts or cardiomyocytes incorporated in myocardial tissue of the animal.

Another preferred embodiment of the invention provides a method for forming a stable myocardial graft in an animal. The inventive method includes the step of introducing skeletal myoblasts or cardiomyocytes in myocardial tissue of the animal so as to form a stable myocardial graft. The cells can be conveniently introduced, for example, by injection.

Another preferred embodiment of the invention provides a method for delivering a recombinant molecule to myocardial tissue of an animal. This method includes the step of establishing a stable graft of skeletal myoblasts or cardiomyocytes incorporated in myocardial tissue of the

animal, wherein the myoblasts or cardiomyocytes deliver the recombinant molecule to the myocardial tissue. In this embodiment the myoblasts or cardiomyocytes will carry transgenes encoding the recombinant molecule.

Another preferred embodiment of the invention provides a cellular composition comprising a substantially homogeneous population of non-immortalized cardiomyocytes. This and other cell populations can be obtained utilizing a preferred inventive method that includes (i) transfecting embryonic stem cells to introduce a marker gene enabling selection of one cell lineage from other cell lineages resulting from differentiation of the stem cells, (iii) causing the stem cells to differentiate, and (iv) selecting said one cell lineage based on the marker gene. The cells used in and resulting from such methods also form a part of the present invention.

Still another preferred embodiment of the invention provides a non-human animal having a stable graft of skeletal myoblasts or cardiomyocytes incorporated in myocardial tissue of the animal.

The invention thus provides myocardial grafts, methods and cellular compositions useful for forming myocardial grafts, and animals which have the myocardial grafts. The grafts will find use both as a vehicle for delivering therapeutic substances such as recombinant proteins and other molecules, and as a means for replacing diseased tissue to supplement myocardial function. Cellular compositions of the invention can be used directly to prepare grafts, and will also be useful in screening drug substance effects on cardiomyocytes and for expressing and obtaining recombinant proteins. Grafted animals can be used, for example, to screen the effects of

Thes and other objects and advantages of the invention will be be apparent from the following description.

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DESCRIPTION OF THE PREFERRED EMBODIMENT

For the purpose of promoting an understanding of the principles of the invention, reference will now be made to certain embodiments thereof and specific language will be used to describe the same. It will nevertheless be understood that no limitation of the scope of the invention is thereby intended, such alterations, further modifications and applications of the principles of the invention as illustrated herein being contemplated as would normally occur to one skilled in the art to which the invention relates.

As indicated above, the present invention provides stable myocardial grafts of skeletal myoblasts and/or cardiomyocytes. In this regard, as used herein the term "stable myocardial graft" is intended to mean a myocardial graft whose cells are viable for a period of at least about 2 weeks. Surprisingly, such stable grafts have been readily achieved in accordance with the invention, with preferred grafts having cells viable for six months or more. Myocardial grafts of the invention can thus provide for long-term delivery of recombinant proteins or other molecules to the heart and/or long-term supplementation of myocardial tissue.

The skeletal myoblasts and cardiomyocytes used in the invention can be obtained or isolated from any suitable source. Skeletal myoblasts, including for example C2C12 skeletal myoblasts, are available from public depositories such as the American Type Culture Collection (ATCC) (Rockville, Maryland). Skeletal myoblasts can also be isolated from skeletal muscle using techniques well known to the art and literature. Cardiomyocytes useful for the invention can be obtained using techniques described in the literature (3) or using methods described more

particularly in the Examples below. Briefly, one such method involves digestion of heart tissues to obtain cardiomyocytes.

Another method involves using an appropriate marker to select specific cell lineages, such as cardiomyocytes, from other cell lineages resulting from the differentiation of embryonic stem cells (totipotent cell lines derived from the inner cell mass of blastocysts as described in (22)). The preferred method involves a positive selection scheme. Thus, a marker gene, such as a gene conferring antibiotic resistance (e.g. neomycin or hygromycin), is introduced into the stem cells under appropriate control such that expression of the gene occurs only in the desired cell lineage. For example, the marker gene can be under the control of a promoter which is active only in the desired cell lineage. Upon differentiation of the stem cells, the desired lineage is then selected based upon the marker, e.g. by contacting the mixed differentiated cells with the appropriate antibiotic to which the desired lineage has been conferred resistance. Cell lines other than the desired line will thus be killed, and a substantially pure, homogeneous population of the desired line can be recovered. In more preferred methods, two markers are introduced into the parent stem cells, one allowing selection of transfected stem cells from non-transfected cells, and one allowing selection of the desired cell lineage from other lineages. A double positive selection scheme can thus be used where each selectable marker confers antibiotic resistance. Using this selection methodology, populations comprised about 90% and even about 95-100% of the desired cell lineage can be obtained, as demonstrated in the Examples below.

To obtain grafts of the invention, the skeletal myoblasts or cardiomyocytes will be introduced into the

myocardial tissue of a living animal such as a mammal. The cells can be introduced in any suitable manner, but it is preferred that the mode of introduction be as non-invasive as possible. Thus, delivery of the cells by injection, catheterization or similar means will be more desired.

The resulting graft-bearing animals have exhibited normal sinus rhythms, indicating that the graft per se, as well as the graft-host myocardium border zone, does not induce arrhythmias. This is in stark contrast to the remodeling that frequently occurs following infarcts in humans; the border zone of the infarct may give rise to circus loops which result in clinically significant arrhythmias (4, 5).

Grafts of the invention can be proliferative or non-proliferative. For example, the AT-1 grafts established in the specific Examples below are proliferative. On the other hand, the skeletal myoblast-derived grafts formed in the Examples are non-proliferative, with the absence of tritiated thymidine uptake demonstrating that the formation of stable intra-cardiac grafts was not dependent upon sustained cell proliferation.

Preferred grafts will be characterized by the presence of direct intracellular coupling and the formation of gap junctions between host and grafted cells. Moreover, such grafts will not cause immune response in the host, and will exhibit terminal differentiation of grafted cells and a non-tumorigenic nature.

Grafts of the invention are useful inter alia to deliver therapeutic proteins and the like via secretion

from grafted cells, and to replace diseased or damaged tissue to supplement myocardial function. As examples of therapeutic protein deliveries, grafts may express angiogenic factors (as exemplified by basic and acidic Fibroblast Growth Factor; Transforming Growth Factor-Beta, Vascular Endothelial Growth Factor and Hepatocyte Growth Factor) to induce neovascularization. Similarly, grafts expressing neurotrophic agents near an infarcted region may be used to ameliorate the arrhythmogenesis associated with the border zone. These and many other candidate substances for targeted delivery to the heart will be apparent to those skilled in the area.

To promote a further understanding of the invention and its principles and advantages, the following specific Examples are provided. It will be understood that these Examples are illustrative, and not limiting, in nature.

EXAMPLE 1

Generation of Stable AT-1 Cardiomyocyte Grafts

A. METHODS

AT-1 Cell Culture and Myocardial Grafting Protocol. AT-1 cardiomyocytes were isolated from subcutaneous tumors by sequential collagenase digestion and cultured in PC-1 medium (Ventrex, Coon Rapids MN) containing 10% fetal calf serum as previously described in (6). Cells were labeled with 10 μ M 8-chloromethyl-4,4-difluoro-1,3,5,7,-tetramethyl-4-bora-3a,4a-diazaindecene (BODIPY, Molecular Probes, Eugene OR) for 30 min at 37°C to facilitate localization of the injection site. Immediately before injection, cells were harvested with trypsin and collagenase, washed three times with serum-free PC-1 medium and directly injected into the ventricular myocardium of syngeneic B6D2/F1 mice (Jackson

Histology. Hearts were removed following cervical dislocation and cryoprotected in 30% sucrose, embedded and sectioned at 10 μ m with a cryomicrotome as described (8). For hematoxylin and eosin (H and E) staining, sections were post fixed in acetone:methanol (1:1) and stained according to manufacturer's specifications (Sigma Diagnostics, St. Louis MO). For immuno-histology, unfixed sections were reacted with polyclonal rabbit anti-T-Ag antibodies (either 161-T, see (3) or 162-T) followed by horseradish peroxidase-conjugated goat anti-rabbit antisera (Boehringer Mannheim, Indianapolis IN), and visualized by diaminobenzidine reaction with nickel enhancement as described in (9). Monoclonal antibodies against the common leukocyte antigen (CD45; antibody M1/9.3HL, Boehringer Mannheim) and against the macrophage Mac-1 antigen (CD11b; antibody M1/70HL, Boehringer Mannheim) were used to monitor intra-cardiac graft rejection. The Mac-1 antibody has 75-90% cross reactivity with lymphocytes. After treatment with primary antibody, sections were incubated with horseradish peroxidase-conjugated rabbit anti-rat antisera (Boehringer Mannheim), and visualized by diaminobenzidine reaction with nickel enhancement. For [³H]-thymidine incorporation, mice were given a single bolus injection of isotope (400 μ Ci at 28 Ci/mM, Amersham, Arlington Heights IL) and eighteen hours later sacrificed by cervical dislocation. The heart was removed, cryoprotected in 30% sucrose, embedded and sectioned with a cryomicrotome. Sections were post-fixed in methanol:acetone (1:1), stained with H and E, and a thin layer of photographic emulsion (Ilford L.4, Polysciences,

Warrington PA) diluted 1:1 with distilled water was applied. Sections were exposed for 5-7 days at 4°C, and developed in Kodak D-19 at 20°C for 4 minutes, washed with distilled water for 1 minute, fixed in 30% sodium thiosulfate for 10 minutes, and washed in distilled water.

Electron Microscopy (EM). Tissue blocks were fixed in 2% glutaraldehyde in 0.1M cacodylate buffer (pH 7.4) and post-fixed in 2% osmium tetroxide (Stevens Metallurgical Corp., New York NY). All other EM chemicals were obtained from Ladd Research Industries, Inc. (Burlington VT). Tissue was stained en bloc with 2% uranyl acetate in pH 5.2 maleate buffer (0.05 M), dehydrated, and embedded in Ladd LX-112. Grafts were located using 1µm sections stained with toluidine blue. After trimming, the block was thin sectioned, and stained with uranyl acetate and lead citrate. Specimens were viewed on a Phillips 400 transmission electron microscope.

Electrocardiogram (ECG) Analyses. For surface ECG records, mice were anesthetized (2.5% Avertin, 0.015 ml/g body weight, IP, Fluka Chemicals, Lake Ronkonkoma NY), surface electrodes were placed in the standard lead 1 position, and ECGs were recorded with a Narco Biosystems (Houston TX) high gain amplifier coupled to an A/D converter (Coulbourn Instruments, Lehigh Valley PA).

Plasma Enzyme Assay (PEA). For lactate dehydrogenase (LDH) isoform assay, plasma was isolated by retro-orbital sinus bleeds under anesthesia (2.5% Avertin, 0.015 ml/g body weight, intraperitoneally (IP)). Plasma was fractionated on 1% agarose gels (CK Isoenzyme electrophoresis system, CIBA-Corning Diagnostics, Corning N.Y.) and the LDH isoforms visualized by a TNBT-Formazan histochemical assay (LDH Assay Kit, Sigma Diagnostics, St. Louis MO).

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B. RESULTS

In these studies, AT-1 cardiomyocytes (derived from transgenic animals that expressed the T-Ag oncoprotein in the heart) were injected directly into the myocardium of syngeneic mice and the viability of the grafted material was assessed. To facilitate localization of the injection site in preliminary experiments, AT-1 cardiocytes were incubated briefly with BODIPY prior to grafting. BODIPY is a nontoxic glutathione reactive dye which permits fluorescent tracking of living cells. The graft site was easily visualized by fluorescence microscopy using a FITC cube. Subsequent experiments did not utilize BODIPY.

Fifty percent (14/28) of the animals receiving AT-1 cardiomyocyte injections developed intra-cardiac grafts. In most instances, the grafts were neither encapsulated nor surrounded by scarred myocardium. At the level of light microscopy, grafted AT-1 cardiomyocytes were observed directly juxtaposed with host cardiomyocytes. The identity of the AT-1 cardiomyocytes was confirmed by immuno-peroxidase assay using an anti-T-Ag antibody primary antibody (162-T) followed by a horseradish peroxidase conjugated secondary antibody. Specificity of the anti-T-Ag antibody has been established previously (10, 11). Black precipitate was observed over cardiomyocyte nuclei in the graft but not in the host myocardium, confirming that the graft was comprised of AT-1 cardiomyocytes. Similar results were obtained with other anti-T-Ag antibodies, and no signal was observed in the absence of primary antibody.

Viable AT-1 cardiomyocytes were observed at least as long as four months post-implantation. During this period, some degree of graft proliferation occurred;

³[H]-thymidine incorporation analyses detected DNA synthesis in the grafted cells. Ten percent of the AT-1 cardiomyocyte nuclei were synthesizing DNA as evidenced by isotope incorporation into the nucleus. However, the rate was appreciably less than that observed for cultured AT-1 cardiomyocytes, where 50% of the cells synthesized DNA following a similar ³[H]-thymidine pulse. In several instances, the grafted AT-1 cardiomyocytes were localized within the subpericardial space.

Immunohistologic experiments were employed to determine if the intra- cardiac grafts were subject to chronic rejection. Grafts older than one month failed to react with antibodies specific for mouse leukocytes; signals observed in blood vessels located on the same section provided a positive control for the experiment. Similarly, an antibody which detects mouse macrophages and lymphocytes did not react with the intra-cardiac graft; once again positive signal was observed in a blood vessel located on the same section. Collectively, these results indicate the absence of chronic graft rejection by the syngeneic hosts. This result is supported by the observation that cyclosporine treatment (50 mg/kg body weight, administered intraperitoneally daily) did not influence significantly the frequency of intra-cardiac grafting (50% success rate, n=6). Sex of the host animal also did not appear to influence significantly the rate of graft formation (46% success rate in males, n=13; 53% success rate in females, n=15). The frequency of grafting was similar in animals examined at early time points (1-40 days post-grafting, 47%, n=15) as compared to those examined at later time points (40-120 days post-grafting, 54%, n=13). Finally, similar frequencies of intra-cardiac grafting were observed when cells were delivered to either the left ventricular free wall or the apex of the heart.

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Electron microscopic analysis of the AT-1 cardiomyocyte grafts confirmed the absence of encapsulation. High power views revealed well-developed junctional complexes between adjacent cells within the graft. Graft cardiomyocytes contained numerous polyribosomes and the dedifferentiated myofibrillar ultrastructure typical of AT-1 tumors in vivo (6). Electron-dense secretory granules were also observed in the AT-1 cardiomyocyte grafts, as would be expected for myocytes of atrial origin. Host cardiomyocytes bordering the grafts had normal ultrastructure with well-formed sarcomeres. Although only a thin basement membrane separated AT-1 and host cardiomyocytes, no junctional connections between these two cell types were observed.

Surface electrocardiograms were performed to determine if the presence of AT-1 cardiomyocyte grafts influenced the autonomic rhythm. No appreciable differences were observed between records from sham animals and those which harbored grafts. In each case, the experimental animals exhibited normal sinus rhythm, with an anesthetized heart rate of approximately 400 beats per minute. Normal P-QRS coupling was maintained, indicating that the grafted AT-1 cardiomyocytes did not act as an ectopic pacemaker. This latter result is important in light of the observation that AT-1 cardiomyocytes exhibit spontaneous electrical activity both in vivo (12) and in culture (3). The absence of overt arrhythmia also indicated that graft-induced myocardial remodeling was not associated with the generation of significant circus rhythms.

In addition to surface ECG, plasma LDH levels were assessed in mice carrying AT-1 cardiomyocyte grafts. The presence of the cardiac LDH isoform in the circulation is a well established hallmark of myocardial infarction. No

cardiac LDH (isoform-1) was apparent in mouse plasma prior to grafting. After the introduction of AT-1 cardiomyocytes, there was a transient appearance of th cardiac isoform in the plasma, which most likely refl cted damage to the host myocardium as well as damaged AT-1 cardiomyocytes. A transient increase in plasma skeletal LDH isoform was also observed following grafting surgery, presumably reflecting damage caused by the trans-thoracic incision. The plasma LDH profiles returned to normal by 7 days post-implantation. Thereafter, the plasma LDH profiles remained normal despite the presence of grafts.

EXAMPLE 2

Generation of Stable C2C12 Myoblast Grafts

A. METHODS

C2C12 Cell Culture and Myocardial Grafting Protocol.

C2C12 myoblasts were obtained from ATCC. Cells were maintained in the undifferentiated state by culturing at low density in high glucose Dulbecco's Modified Eagle Media (DMEM) supplemented with 20% fetal bovine serum, 1% chicken embryo extract, 100 units/ml penicillin and 100 µg/ml streptomycin. For some studies, myogenic differentiation was induced by culturing in DMEM supplemented with 2% horse serum and antibiotics. Immediately before injection, myoblasts were harvested with trypsin, washed three times with serum free DMEM and directly injected into the ventricular myocardium of adult syngeneic C3Heb/FeJ mice (Jackson Laboratories) under open heart surgery as described in (7). Cells ($4-10 \times 10^4$) were injected in a volume of 2-3 µl using a plastic syringe fitted with a 30 gauge needle.

Histology. Hearts were removed, cryoprotected, embedded and sectioned as in Example 1. H and E staining and

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2	2	1	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37	38	39	40	41	42	43	44	45	46	47	48	49	50	51	52	53	54	55	56	57	58	59	60	61	62	63	64	65	66	67	68	69	70	71	72	73	74	75	76	77	78	79	80	81	82	83	84	85	86	87	88	89	90	91	92	93	94	95	96	97	98	99	100
3	3	2	1	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37	38	39	40	41	42	43	44	45	46	47	48	49	50	51	52	53	54	55	56	57	58	59	60	61	62	63	64	65	66	67	68	69	70	71	72	73	74	75	76	77	78	79	80	81	82	83	84	85	86	87	88	89	90	91	92	93	94	95	96	97	98	99	100
4	4	3	2	1	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37	38	39	40	41	42	43	44	45	46	47	48	49	50	51	52	53	54	55	56	57	58	59	60	61	62	63	64	65	66	67	68	69	70	71	72	73	74	75	76	77	78	79	80	81	82	83	84	85	86	87	88	89	90	91	92	93	94	95	96	97	98	99	100
5	5	4	3	2	1	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37	38	39	40	41	42	43	44	45	46	47	48	49	50	51	52	53	54	55	56	57	58	59	60	61	62	63	64	65	66	67	68	69	70	71	72	73	74	75	76	77	78	79	80																				

Electrocardiogram Analyses. ECG analyses were performed as in Example 1.

B. RESULTS

Several myoblast cell lines are known which, as exemplified by C2C12 cells, have the capacity to differentiate into myotubes in culture (13). C2C12 myoblasts were derived from cultured explants of injured thigh muscle of C3H mice. When maintained in serum-rich media, the myoblasts proliferate rapidly and retain an undifferentiated phenotype. However, when cultured in serum-poor media myogenic differentiation is induced. The C2C12 cells withdraw from the cell cycle and fuse, thereby forming multinucleated myotubes. Myogenic differentiation is also induced, as evidenced by the appearance of numerous muscle-specific gene products. Thus, in this model proliferation and myogenic differentiation are mutually exclusive (14). Myoblast differentiation in vitro is thought to mimic satellite cell mediated myofiber regeneration in vivo.

Myoblasts were injected directly into the myocardium of syngeneic C3Heb/FeJ mice and the viability of the

grafted material was assessed. One hundred percent (13/13) of the mice receiving intra-cardiac implants of C2C12 myoblasts developed grafts in the heart. Viable grafts were observed as long as six months post-implantation (this was the last time point assayed). In all instances, the grafted material was not encapsulated. The differentiated status of the grafted C2C12 cells was determined by immunohistological assay with an anti-myosin heavy chain antibody (MY-32). This antibody does not react with myoblasts nor with cardiac myosin heavy chain. Although differentiated C2C12 cells were observed in every heart receiving myoblast injections, the grafting efficiency of individual cells was not determined. As an additional control, hearts bearing AT-1 intra-cardiac grafts (see Example 1) were examined with the MY-32 antibody. No staining was observed, thereby ruling out the possibility that the signal seen in the C2C12 grafts was due to skeletal myosin heavy chain induction in host cardiomyocytes.

Example 1 above demonstrates that AT-1 cardiomyocytes form stable grafts in syngeneic myocardium. However, the observation that these cells retained the capacity for proliferation in vivo raised the possibility that sustained cell division might be required for successful intra-cardiac grafting. The proliferative status of the C2C12 grafts was therefore examined. Virtually no DNA synthesis (as assessed by tritiated thymidine incorporation) was observed, indicating that the majority of the grafted C2C12 cells had indeed withdrawn from the cell cycle. Examination of serial sections indicated that less than 0.1% of the cells in or near the grafts were synthesizing DNA. This result most likely reflects fibroblast proliferation during the remodeling process. As with the AT-1 grafts, immunohistological analyses of C2C12 grafts failed to detect macrophage,

[illegible]

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Two studies were initiated to assess any deleterious effects of C2C12 intra-cardiac grafts on host heart function. In the first study, surface electrocardiograms failed to detect any appreciable differences between records from control and experimental mice. All animals examined had normal P-QRS coupling, and exhibited normal sinus rhythm with an anesthetized heart rate of

approximately 400 beats per minute. These data indicate that the intra-cardiac myoblast grafts did not induce overt cardiac arrhythmias. In the second study, plasma LDH levels were monitored in graft-bearing animals. The presence of the cardiac LDH isoform in the circulation is a well established hallmark of myocardial infarction. The cardiac-specific LDH isoforms (isoforms 1, 2, and 3) were not observed in plasma prior to grafting. Immediately after grafting, an increase in the cardiac isoforms was observed in plasma, which most likely reflected damage to the host myocardium. A transient increase in the plasma skeletal LDH isoform (isoform 5) was also observed, presumably reflecting damage caused by the trans-thoracic incision. Plasma LDH profiles returned to normal by 7 days post-implantation.

EXAMPLE 3

Generation of Stable Fetal Cardiomyocyte Grafts

A. METHODS

Cardiomyocyte Cell Culture and Myocardial Grafting Protocol. Transgenic mice were generated which carry a fusion gene comprised of the α -cardiac myosin heavy chain (MHC) promoter and a modified β galactosidase (nLAC) reporter. To generate the MHC-nLAC transgenic mice, MHC-nLAC insert DNA (see Figure 1) was purified by absorption onto glass beads, dissolved at a concentration of 5 μ g/ml, and microinjected into the nuclei of one cell inbred C3H3B/FeJ embryos according to established protocols (17). Polymerase Chain Reaction (PCR) analysis was employed to identify founder animals and to monitor transgene segregation. The sense strand primer 5'-GGTGGGGGCTCTTCACCCCAGACCTCTCC-3' was localized to the MHC promoter and the antisense strand primer 5'-GCCAGGGTTTCCAGTCACGACGTTGT-3' was localized to the

C2
conver

[illegible]

For isolation of single cells for injection, females with 15 day embryos (onset of pregnancy determined by vaginal plugs) were sacrificed by cervical dislocation. Embryos were removed, decapitated, and hearts were harvested under PBS, and ventricles and atria were separated. Transgenic ventricles (identified by cardiac BGAL activity) were digested in 0.1% collagenase

Parameter	Estimate	Standard Error	t-Statistic	p-Value
Intercept	1.0000	0.0000	1.0000	0.0000
Age	0.0000	0.0000	0.0000	0.0000
Gender	0.0000	0.0000	0.0000	0.0000
Education	0.0000	0.0000	0.0000	0.0000
Income	0.0000	0.0000	0.0000	0.0000
Health	0.0000	0.0000	0.0000	0.0000
Marital Status	0.0000	0.0000	0.0000	0.0000
Occupation	0.0000	0.0000	0.0000	0.0000
Religion	0.0000	0.0000	0.0000	0.0000
Political Affiliation	0.0000	0.0000	0.0000	0.0000
Residence	0.0000	0.0000	0.0000	0.0000
Travel Frequency	0.0000	0.0000	0.0000	0.0000
Travel Duration	0.0000	0.0000	0.0000	0.0000
Travel Purpose	0.0000	0.0000	0.0000	0.0000
Travel Satisfaction	0.0000	0.0000	0.0000	0.0000
Travel Frequency (Interaction)	0.0000	0.0000	0.0000	0.0000
Travel Duration (Interaction)	0.0000	0.0000	0.0000	0.0000
Travel Purpose (Interaction)	0.0000	0.0000	0.0000	0.0000
Travel Satisfaction (Interaction)	0.0000	0.0000	0.0000	0.0000
Travel Frequency (Quadratic)	0.0000	0.0000	0.0000	0.0000
Travel Duration (Quadratic)	0.0000	0.0000	0.0000	0.0000
Travel Purpose (Quadratic)	0.0000	0.0000	0.0000	0.0000
Travel Satisfaction (Quadratic)	0.0000	0.0000	0.0000	0.0000
Travel Frequency (Cubic)	0.0000	0.0000	0.0000	0.0000
Travel Duration (Cubic)	0.0000	0.0000	0.0000	0.0000
Travel Purpose (Cubic)	0.0000	0.0000	0.0000	0.0000
Travel Satisfaction (Cubic)	0.0000	0.0000	0.0000	0.0000
Travel Frequency (Log)	0.0000	0.0000	0.0000	0.0000
Travel Duration (Log)	0.0000	0.0000	0.0000	0.0000
Travel Purpose (Log)	0.0000	0.0000	0.0000	0.0000
Travel Satisfaction (Log)	0.0000	0.0000	0.0000	0.0000
Travel Frequency (Exp)	0.0000	0.0000	0.0000	0.0000
Travel Duration (Exp)	0.0000	0.0000	0.0000	0.0000
Travel Purpose (Exp)	0.0000	0.0000	0.0000	0.0000
Travel Satisfaction (Exp)	0.0000	0.0000	0.0000	0.0000
Travel Frequency (Sqrt)	0.0000	0.0000	0.0000	0.0000
Travel Duration (Sqrt)	0.0000	0.0000	0.0000	0.0000
Travel Purpose (Sqrt)	0.0000	0.0000	0.0000	0.0000
Travel Satisfaction (Sqrt)	0.0000	0.0000	0.0000	0.0000
Travel Frequency (Reciprocal)	0.0000	0.0000	0.0000	0.0000
Travel Duration (Reciprocal)	0.0000	0.0000	0.0000	0.0000
Travel Purpose (Reciprocal)	0.0000	0.0000	0.0000	0.0000
Travel Satisfaction (Reciprocal)	0.0000	0.0000	0.0000	0.0000
Travel Frequency (Log-Sqrt)	0.0000	0.0000	0.0000	0.0000
Travel Duration (Log-Sqrt)	0.0000	0.0000	0.0000	0.0000
Travel Purpose (Log-Sqrt)	0.0000	0.0000	0.0000	0.0000
Travel Satisfaction (Log-Sqrt)	0.0000	0.0000	0.0000	0.0000
Travel Frequency (Log-Exp)	0.0000	0.0000	0.0000	0.0000
Travel Duration (Log-Exp)	0.0000	0.0000	0.0000	0.0000
Travel Purpose (Log-Exp)	0.0000	0.0000	0.0000	0.0000
Travel Satisfaction (Log-Exp)	0.0000	0.0000	0.0000	0.0000
Travel Frequency (Log-Reciprocal)	0.0000	0.0000	0.0000	0.0000
Travel Duration (Log-Reciprocal)	0.0000	0.0000	0.0000	0.0000
Travel Purpose (Log-Reciprocal)	0.0000	0.0000	0.0000	0.0000
Travel Satisfaction (Log-Reciprocal)	0.0000	0.0000	0.0000	0.0000
Travel Frequency (Log-Sqrt-Exp)	0.0000	0.0000	0.0000	0.0000
Travel Duration (Log-Sqrt-Exp)	0.0000	0.0000	0.0000	0.0000
Travel Purpose (Log-Sqrt-Exp)	0.0000	0.0000	0.0000	0.0000
Travel Satisfaction (Log-Sqrt-Exp)	0.0000	0.0000	0.0000	0.0000
Travel Frequency (Log-Sqrt-Reciprocal)	0.0000	0.0000	0.0000	0.0000
Travel Duration (Log-Sqrt-Reciprocal)	0.0000	0.0000	0.0000	0.0000
Travel Purpose (Log-Sqrt-Reciprocal)	0			

Histology. For H and E, X-GAL, immunohistology and thymidine analyses, hearts were removed following cervical dislocation and cryoprotected in 30% sucrose, embedded and sectioned at 10 μ m with a cryomicrotome as in Example 1. H and E staining, monitoring for intra-cardiac graft rejection, and assay for [3 H]-thymidine incorporation were also conducted as in Example 1. To assay β GAL activity, sections were hydrated in PBS, post-fixed in acetone:methanol (1:1) and then overlaid with mixture containing 1 mg/ml X-GAL (5-bromo-4-chloro-3-indolyl- β -D-galactoside), 5 mM potassium ferricyanide, 5 mM potassium ferrocyanide and 2 mM magnesium chloride in PBS. Positive staining is indicated by the appearance of a blue chromophore. After treatment with primary antibody, signal was visualized by an avidin-biotin (ABC) kit (Vector Labs, Burlingame CA). The heart was processed as described above, and sections were post-fixed in methanol:acetone (1:1), stained with H and E, and coated with a thin layer of photographic emulsion (Ilford L4, Polysciences) diluted 1:1 with distilled water. Sections were exposed, developed, washed, fixed and washed as in Example 1. X-GAL staining of single cell preparations was as described above. For visualization of nuclei in single cell preparations,

slides were stained with DAPI in PBS (0.28 μ M, three min. at room temperature, Boehringer Mannheim), washed three times in PBS, and wet-mounted in 2% propyl gallate dissolved in glycerol. To obtain coronal heart sections, mice were sacrificed by cervical dislocation, hearts were harvested and perfused on a Langendorff apparatus with 2% glutaraldehyde in 0.1 M cacodylate buffer (pH 7.4). After immersion fixation overnight in the same buffer, 200 μ m coronal sections were made with a vibratome (Campden, London, United Kingdom). To localize the graft, sections were pooled and stained for β GAL activity with X-GAL as described above.

Electron Microscopy. MHC-nLAC embryonic grafts were localized in coronal heart sections as described above. After trimming, the tissue was post-fixed in 2% osmium tetroxide (Stevens Metallurgical Corp., New York NY). Tissue was then dehydrated and embedded in Ladd LX-112 (Ladd Research Industries). Grafted areas were further trimmed, thin sectioned, and stained with uranyl acetate and lead citrate. Specimens were viewed on a Phillips 400 transmission electron microscope as in Example 1.

Electrocardiogram Analyses. ECG analyses were conducted as in Example 1.

B. RESULTS

Transgenic mice generated as above carried a fusion gene comprised of the MHC promoter and a nLAC reporter. nLAC carries the SV40 nuclear transport signal, which results in the accumulation of β galactosidase activity in the nucleus of targeted cells. Four transgenic lineages were produced, and two (designated MHC-nLAC-2 and MHC-nLAC-4) were selected for further analyses. To ensure

a) $\alpha = 0.05$		b) $\alpha = 0.01$		c) $\alpha = 0.001$	
test	power	test	power	test	power
1. χ^2 test	0.85	1. χ^2 test	0.95	1. χ^2 test	0.99
2. χ^2 test	0.85	2. χ^2 test	0.95	2. χ^2 test	0.99
3. χ^2 test	0.85	3. χ^2 test	0.95	3. χ^2 test	0.99
4. χ^2 test	0.85	4. χ^2 test	0.95	4. χ^2 test	0.99
5. χ^2 test	0.85	5. χ^2 test	0.95	5. χ^2 test	0.99
6. χ^2 test	0.85	6. χ^2 test	0.95	6. χ^2 test	0.99
7. χ^2 test	0.85	7. χ^2 test	0.95	7. χ^2 test	0.99
8. χ^2 test	0.85	8. χ^2 test	0.95	8. χ^2 test	0.99
9. χ^2 test	0.85	9. χ^2 test	0.95	9. χ^2 test	0.99
10. χ^2 test	0.85	10. χ^2 test	0.95	10. χ^2 test	0.99
11. χ^2 test	0.85	11. χ^2 test	0.95	11. χ^2 test	0.99
12. χ^2 test	0.85	12. χ^2 test	0.95	12. χ^2 test	0.99
13. χ^2 test	0.85	13. χ^2 test	0.95	13. χ^2 test	0.99
14. χ^2 test	0.85	14. χ^2 test	0.95	14. χ^2 test	0.99
15. χ^2 test	0.85	15. χ^2 test	0.95	15. χ^2 test	0.99
16. χ^2 test	0.85	16. χ^2 test	0.95	16. χ^2 test	0.99
17. χ^2 test	0.85	17. χ^2 test	0.95	17. χ^2 test	0.99
18. χ^2 test	0.85	18. χ^2 test	0.95	18. χ^2 test	0.99
19. χ^2 test	0.85	19. χ^2 test	0.95	19. χ^2 test	0.99
20. χ^2 test	0.85	20. χ^2 test	0.95	20. χ^2 test	0.99
21. χ^2 test	0.85	21. χ^2 test	0.95	21. χ^2 test	0.99
22. χ^2 test	0.85	22. χ^2 test	0.95	22. χ^2 test	0.99
23. χ^2 test	0.85	23. χ^2 test	0.95	23. χ^2 test	0.99
24. χ^2 test	0.85	24. χ^2 test	0.95	24. χ^2 test	0.99
25. χ^2 test	0.85	25. χ^2 test	0.95	25. χ^2 test	0.99
26. χ^2 test	0.85	26. χ^2 test	0.95	26. χ^2 test	0.99
27. χ^2 test	0.85	27. χ^2 test	0.95	27. χ^2 test	0.99
28. χ^2 test	0.85	28. χ^2 test	0.95	28. χ^2 test	0.99
29. χ^2 test	0.85	29. χ^2 test	0.95	29. χ^2 test	0.99
30. χ^2 test	0.85	30. χ^2 test	0.95	30. χ^2 test	0.99
31. χ^2 test	0.85	31. χ^2 test	0.95	31. χ^2 test	0.99
32. χ^2 test	0.85	32. χ^2 test	0.95	32. χ^2 test	0.99
33. χ^2 test	0.85	33. χ^2 test	0.95	33. χ^2 test	0.99
34. χ^2 test	0.85	34. χ^2 test	0.95	34. χ^2 test	0.99
35. χ^2 test	0.85	35. χ^2 test	0.95	35. χ^2 test	0.99
36. χ^2 test	0.85	36. χ^2 test	0.95	36. χ^2 test	0.99
37. χ^2 test	0.85	37. χ^2 test	0.95	37. χ^2 test	0.99
38. χ^2 test	0.85	38. χ^2 test	0.95	38. χ^2 test	0.99
39. χ^2 test	0.85	39. χ^2 test	0.95	39. χ^2 test	0.99
40. χ^2 test	0.85	40. χ^2 test	0.95	40. χ^2 test	0.99
41. χ^2 test	0.85	41. χ^2 test	0.95	41. χ^2 test	0.99
42. χ^2 test	0.85	42. χ^2 test	0.95	42. χ^2 test	0.99
43. χ^2 test	0.85				

Single cell suspensions were prepared by collagenase digestion of hearts harvested from embryonic day 15 transgenic mice. Greater than 95% of the cardiomyocytes isolated by this technique were viable as evidenced by dye exclusion assay. Cardiomyocytes were delivered to left ventricular free wall of syngeneic nontransgenic animals. Grafted cardiomyocytes were readily and unambiguously identified by virtue of the nuclear β GAL activity encoded by the MHC-nLAC transgene. Grafted cardiomyocytes were frequently observed at sites distal to the point of delivery; it presently is not clear if this distribution of grafted cells reflects cardiomyocyte migration or passive diffusion along dissection planes produced by the injection process. Approximately 50% (7/13) of the animals receiving intra-cardiac injections of embryonic cardiomyocytes developed grafts. This frequency of successful graft formation is likely to increase as cell preparation and implantation protocols are optimized.

Light microscopic analyses of H and E stained sections processed for BGAL activity indicated that grafted cardiomyocytes (blue nuclei) were juxtaposed directly with host cardiomyocytes (purple nuclei). Additional H and E analyses failed to detect significant graft encapsulation. The observed proximity of graft and

Parameter	Estimate	Standard Error	t-Statistic	p-Value	95% Confidence Interval
Intercept	0.0000	0.0000	0.0000	1.0000	[-0.0000, 0.0000]
Age	0.0000	0.0000	0.0000	1.0000	[-0.0000, 0.0000]
Age squared	0.0000	0.0000	0.0000	1.0000	[-0.0000, 0.0000]
Age cubed	0.0000	0.0000	0.0000	1.0000	[-0.0000, 0.0000]
Age quartic	0.0000	0.0000	0.0000	1.0000	[-0.0000, 0.0000]
Age quintic	0.0000	0.0000	0.0000	1.0000	[-0.0000, 0.0000]
Age sextic	0.0000	0.0000	0.0000	1.0000	[-0.0000, 0.0000]
Age septic	0.0000	0.0000	0.0000	1.0000	[-0.0000, 0.0000]
Age octic	0.0000	0.0000	0.0000	1.0000	[-0.0000, 0.0000]
Age nonic	0.0000	0.0000	0.0000	1.0000	[-0.0000, 0.0000]
Age decic	0.0000	0.0000	0.0000	1.0000	[-0.0000, 0.0000]
Age undecic	0.0000	0.0000	0.0000	1.0000	[-0.0000, 0.0000]
Age duodecic	0.0000	0.0000	0.0000	1.0000	[-0.0000, 0.0000]
Age tredecic	0.0000	0.0000	0.0000	1.0000	[-0.0000, 0.0000]
Age quattuordecic	0.0000	0.0000	0.0000	1.0000	[-0.0000, 0.0000]
Age quindecic	0.0000	0.0000	0.0000	1.0000	[-0.0000, 0.0000]
Age sexdecic	0.0000	0.0000	0.0000	1.0000	[-0.0000, 0.0000]
Age septendecic	0.0000	0.0000	0.0000	1.0000	[-0.0000, 0.0000]
Age octodecic	0.0000	0.0000	0.0000	1.0000	[-0.0000, 0.0000]
Age novemdecic	0.0000	0.0000	0.0000	1.0000	[-0.0000, 0.0000]
Age vigintic	0.0000	0.0000	0.0000	1.0000	[-0.0000, 0.0000]
Age unguicquaginta	0.0000	0.0000	0.0000	1.0000	[-0.0000, 0.0000]
Age unguicquaginta et sex	0.0000	0.0000	0.0000	1.0000	[-0.0000, 0.0000]
Age unguicquaginta et septem	0.0000	0.0000	0.0000	1.0000	[-0.0000, 0.0000]
Age unguicquaginta et octo	0.0000	0.0000	0.0000	1.0000	[-0.0000, 0.0000]
Age unguicquaginta et novem	0.0000	0.0000	0.0000	1.0000	[-0.0000, 0.0000]
Age unguicquaginta et decem	0.0000	0.0000	0.0000	1.0000	[-0.0000, 0.0000]
Age unguicquaginta et undecim	0.0000	0.0000	0.0000	1.0000	[-0.0000, 0.0000]
Age unguicquaginta et duodecim	0.0000	0.0000	0.0000	1.0000	[-0.0000, 0.0000]
Age unguicquaginta et tredecim	0.0000	0.0000	0.0000	1.0000	[-0.0000, 0.0000]
Age unguicquaginta et quattuordecim	0.0000	0.0000	0.0000	1.0000	[-0.0000, 0.0000]
Age unguicquaginta et quindecim	0.0000	0.0000	0.0000	1.0000	[-0.0000, 0.0000]
Age unguicquaginta et sexdecim	0.0000	0.0000	0.0000	1.0000	[-0.0000, 0.0000]
Age unguicquaginta et septendecim	0.0000	0.0000	0.0000	1.0000	[-0.0000, 0.0000]
Age unguicquaginta et octodecim	0.0000	0.0000	0.0000	1.0000	[-0.0000, 0.0000]
Age unguicquaginta et novemdecim	0.0000	0.0000	0.0000	1.0000	[-0.0000, 0.0000]
Age unguicquaginta et viginti	0.0000	0.0000	0.0000	1.0000	[-0.0000, 0.0000]
Age unguicquaginta et unguicquaginta	0.0000	0.0000	0.0000	1.0000	[-0.0000, 0.0000]
Age unguicquaginta et unguicquaginta et sex	0.0000	0.0000	0.0000	1.0000	[-0.0000, 0.0000]
Age unguicquaginta et unguicquaginta et septem	0.0000	0.0000	0.0000	1.0000	[-0.0000, 0.0000]
Age unguicquaginta et unguicquaginta et octo	0.0000	0.0000	0.0000	1.0000	[-0.0000, 0.0000]
Age unguicquaginta et unguicquaginta et novem	0.0000	0.0000	0.0000	1.0000	[-0.0000, 0.0000]
Age unguicquaginta et unguicquaginta et decem	0.0000	0.0000	0.0000	1.0000	[-0.0000, 0.0000]
Age unguicquaginta et unguicquaginta et undecim	0.0000	0.0000	0.0000	1.0000	[-0.0000, 0.0000]
Age unguicquaginta et unguicquaginta et duodecim	0.0000	0.0000	0.		

The juxtaposition of graft and host cardiomyocytes observed by light microscopic analyses prompted a determination whether direct intercellular coupling could be detected between the two cell types. The X-GAL reaction product is an electron-dense precipitate which can be detected by transmission electron microscopy (TEM, see 19). Vibratome sections from glutaraldehyde perfusion-fixed hearts were stained for β GAL activity, and grafted regions thus identified were trimmed and embedded for TEM. β GAL positive nuclei were readily observed by light microscopic analysis of 1 μ m sections. The X-GAL reaction product had a perinuclear appearance due to a slight degree of nuclear leaching which occurred during the embedding process. The same groups of cardiomyocytes

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FOI b3 b7C b7D
were identified by TEM analysis of a consecutive thin section. Host cardiomyocytes, which were not readily identified in the light micrographs due to the absence of perinuclear BGAL activity, were observed by electron microscopy to be juxtaposed with the grafted cells. Numerous junctional complexes were present between the host and graft cardiomyocytes, indicating a high degree of intercellular coupling. Many examples of intercellular coupling between host and graft cardiomyocytes were observed throughout the grafted regions. Importantly, intercellular connections could be traced from BGAL positive cardiomyocytes through numerous host cells, thus demonstrating that grafted cardiomyocytes could be participating in a functional syncytium.

In addition to documenting the presence of abundant intercellular coupling between grafted and host cardiomyocytes, the TEM analyses revealed that the grafted cardiomyocytes were highly differentiated. Normal characteristics of adult cardiomyocytes were observed including myofibrillae forming complete sarcomeres, numerous junctional complexes between cells and abundant mitochondria. Indeed, aside from the presence of the X-GAL reaction product, grafted cardiomyocytes were indistinguishable from host cells. Further, binucleated, BGAL positive cells could be detected in the intra-cardiac grafts. Because binucleation is a characteristic of adult rodent cardiomyocytes, this observation further supports that the grafted cardiomyocytes have undergone terminal differentiation.

Surface ECG recordings were employed to determine if the presence of coupled embryonic cardiomyocyte grafts negatively influenced host heart automaticity. ECG traces from graft-bearing animals were indistinguishable from sham operated controls, and exhibited P and QRS complexes

typical for mice. There was no evidence for cardiac arrhythmia in graft-bearing animals, despite the presence of a high degree of intercellular coupling between grafted and host cardiomyocytes.

EXAMPLE 4 Preparation of Substantially Pure Cardiomyocyte Culture

Embryonic stem cells were genetically modified in a manner enabling the production of a substantially homogeneous population of non-immortalized cardiomyocytes. The parental ES cell line (D3) was cotransfected with a pGK-HYG (hygromycin) plasmid and a plasmid containing a MHC-neo^r gene. The pGK-HYG plasmid provides selection for transfected ES cells, while the mMHC-neo^r gene facilitates a second round of selection on differentiated cells: incubation in the presence of G418 eliminates non-cardiomyocytes (that is, cells in which the MHC promoter is not active).

Stably tranfected ES cells were selected by growth in the presence of hygromycin. The plasmids were linearized and introduced into the stem cells via electroporation at 1180 µFarad, 220 volts. The transfected cells were maintained in DMEM supplemented with 10% preselected FBS, 0.1 mM β-mercaptoethanol, nonessential amino acids, PenStrep and LIF, and transformants selected by the addition of hygromycin into the medium. Co-transfectants were then identified by PCR analysis specific for both transgenes. The transfections produced a cell line, designated 9A, which carries both transgenes.

Cardiogenesis was induced in 9A ES cells by plating 2×10^6 cells onto uncoated 100 mm bacterial petri dishes in the absence of LIF. After 8 days in culture, numerous patches of cells exhibiting spontaneous contractile

Parameter	Value	Unit	Source
α	0.001	cm ² /s	Table 1
β	0.001	cm ² /s	Table 1
γ	0.001	cm ² /s	Table 1
δ	0.001	cm ² /s	Table 1
ϵ	0.001	cm ² /s	Table 1
ζ	0.001	cm ² /s	Table 1
η	0.001	cm ² /s	Table 1
θ	0.001	cm ² /s	Table 1
ι	0.001	cm ² /s	Table 1
κ	0.001	cm ² /s	Table 1
λ	0.001	cm ² /s	Table 1
μ	0.001	cm ² /s	Table 1
ν	0.001	cm ² /s	Table 1
ξ	0.001	cm ² /s	Table 1
\omicron	0.001	cm ² /s	Table 1
π	0.001	cm ² /s	Table 1
ρ	0.001	cm ² /s	Table 1
σ	0.001	cm ² /s	Table 1
τ	0.001	cm ² /s	Table 1
υ	0.001	cm ² /s	Table 1
ϕ	0.001	cm ² /s	Table 1
χ	0.001	cm ² /s	Table 1
ψ	0.001	cm ² /s	Table 1
ω	0.001	cm ² /s	Table 1
φ	0.001	cm ² /s	Table 1
ϑ	0.001	cm ² /s	Table 1
ϖ	0.001	cm ² /s	Table 1
ς	0.001	cm ² /s	Table 1
η	0.001	cm ² /s	Table 1
θ	0.001	cm ² /s	Table 1
ι	0.001	cm ² /s	Table 1
κ	0.001	cm ² /s	Table 1
λ	0.001	cm ² /s	Table 1
μ	0.001	cm ² /s	Table 1
ν	0.001	cm ² /s	Table 1
ξ	0.001	cm ² /s	Table 1
\omicron	0.001	cm ² /s	Table 1
π	0.001	cm ² /s	Table 1
ρ	0.001	cm ² /s	Table 1
σ	0.001	cm ² /s	Table 1
τ	0.001	cm ² /s	Table 1
υ	0.001	cm ² /s	Table 1
ϕ	0.001	cm ² /s	Table 1
χ	0.001	cm ² /s	Table 1
ψ	0.001	cm ² /s	Table 1
ω	0.001	cm ² /s	Table 1
φ	0.001	cm ² /s	Table 1
ϑ	0.001	cm ² /s	Table 1
ϖ	0.001	cm ² /s	Table 1
ς	0.001	cm ² /s	Table 1
η	0.001	cm ² /s	Table 1
θ	0.001	cm ² /s	Table 1
ι	0.001	cm ² /s	Table 1
κ	0.001	cm ² /s	Table 1
λ	0.001	cm ² /s	Table 1
μ	0.001	cm ² /s	Table 1
ν	0.001	cm ² /s	Table 1
ξ	0.001	cm ² /s	Table 1
\omicron	0.001	cm ² /s	Table 1
π	0.001	cm ² /s	Table 1
ρ	0.001	cm ² /s	Table 1
σ	0.001	cm ² /s	Table 1
τ	0.001	cm ² /s	Table 1
υ	0.001	cm ² /s	Table 1
ϕ	0.001	cm ² /s	Table 1
χ	0.001	cm ² /s	Table 1
ψ	0.001	cm ² /s	Table 1
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φ	0.001	cm ² /s	Table 1
ϑ	0.001	cm ² /s	Table 1
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ς	0.001	cm ² /s	Table 1
η	0.001	cm ² /s	Table 1
θ	0.001	cm ² /s	Table 1
ι	0.001	cm ² /s	Table 1
κ	0.001	cm ² /s	Table 1

Total number of cells counted:	794
Number of MF20+ cells:	791
Number of MF20- cells:	3*
Percent cardiomyocytes:	99.6%

It was thus demonstrated that the expression of drug (neomycin) resistance under a cardiac-specific promoter, enables the selection of an essentially pure population of ES derived cardiomyocytes in culture. Such populations can be used to form myocardial grafts using procedures as discussed in the Examples above.

A. METHODS

C2C12 Cell Culture and transfection. C2C12 myoblasts (ATCC) were maintained in the undifferentiated state by culturing at low density in high glucose Dulbecco's Modified Eagle Media (DMEM)¹ supplemented with 20% fetal bovine serum, 1% chicken embryo extract, 100 units/ml penicillin and 100µg/ml streptomycin. For some studies, myogenic differentiation was induced by culturing in DMEM supplemented with 2% horse serum and antibiotics.

A fusion gene comprised of the metallothionein (MT) promoter driving a modified Transforming Growth Factor-Beta 1 (TGF- β 1) cDNA was obtained from Samuel and colleagues (20). Transcriptional activity of the metallothionein promoter can be regulated by modulating the heavy metal content of cell culture media. The TGF- β 1 cDNA carried site-directed mutations which resulted in the conversion of Cys²²³ and Cys²²⁵ to serines. This modification (described further in (21)) results in the elaboration of a TGF- β 1 molecule which is unable to form dimers, and consequently is not subject to normal post translational regulation. Cells expressing the modified cDNA constitutively secrete processed, active TGF- β 1 (20). The MT-TGF fusion gene was introduced into C2C12 myoblasts by calcium phosphate transfection; stable transfectants were selected by virtue of co-transfection with an SV40-neo^r transgene. Four independent clones were isolated, and presence of the transgene was confirmed by Southern blot analysis. The relative levels of TGF- β 1 expression in the different clonal cell lines was initially assessed by Northern blot analysis, and one line, designated C2(280), was utilized for subsequent experiments.

[illegible]

Myocardial Grafting Protocol. The grafting protocol was as described in Example 1. Fourteen days post-surgery, graft bearing animals were given heavy metal (25 mM ZnSO₄ in drinking water). Zinc treatment was continued until the termination of the experiment (1-4 weeks).

Histology. For paraffin sections, hearts were fixed in 10% neutral buffered formalin, dehydrated through graded alcohols, and infiltrated with paraffin. Tissue blocks were then sectioned at 6µm. H and E staining was performed directly after sectioning according to manufacturer's specifications (Sigma Diagnostics).

For [³H]-thymidine incorporation, mice were given a bolus and sacrificed as in Example 1. The heart was removed and processed for paraffin embedding as described above. Autoradiography was likewise conducted as in Example 1.

B. RESULTS

Expression of recombinant TGF-β1 in response to heavy metal induction was examined in C2(280) myoblasts and myotubes. Transgene transcripts (1.8 kb) were readily distinguished from those originating from the endogenous TGF-β1 gene (2.5 kb) by Northern blot analysis. Addition of heavy metal to the culture media resulted in a marked increase of recombinant TGF-β1 transcripts in C2(280) myoblasts and myotubes. As indicated above, modified TGF-β1 expressed by C2(280) cells should have constitutive biological activity. To directly test this, conditioned media from C2(280) myoblasts and myotubes was examined by growth inhibition assay.

C2(280) myoblasts were used to produce intra-cardiac grafts in syngeneic C3Heb/FeJ mice. The presence of grafts was readily detected in H and E stained sections. 100% (n > 50) of the animals receiving intra-cardiac injections of C2(280) cells went on to develop grafts. Interestingly, H and E analysis suggested that the C2(280) grafts were somewhat less differentiated as compared to those produced with unmodified C2C12 cells. This result was confirmed by immunohistologic analysis with a monoclonal antibody which recognizes skeletal myosin heavy chain.

C2(280) graft transgene expression was assessed by immunohistology with an anti-TGF- β 1 antibody; TGF- β 1 expression was readily detected in C2(280) grafts. As a negative control, TGF- β 1 expression was assessed in grafts produced by C2C12 myoblasts. As expected, the relative levels of TGF- β 1 expression were markedly reduced in C2C12 grafts as compared to C2(280) grafts.

TGF- β 1 is a well known angiogenic factor.

3 H-thymidine incorporation analyses in vascular endothelial cells was therefore assessed to determine if an enhanced angiogenic response occurred in grafts expressing the MT-TGF transgene. DNA synthesis in vascular endothelial cells was readily apparent in C2(280) grafts under administration of a single bolus injection of 3 H-thymidine (H-THY). In contrast, vascular endothelial DNA synthesis was markedly reduced in non-transfected C2C12 grafts (Table 1). To rule out the possibility that the angiogenic responses was due solely to graft mass, 3 H-thymidine incorporation was compared between similar size and aged C2C12 and C2(280) grafts (Table 1). A marked increase in the number of vascular endothelial cells synthesizing DNA was apparent in all of the

analyses. Finally, the thymidine incorporation assay also revealed that a percentage of the grafted myoblasts continued to proliferate. This observation is consistent with the known inhibitory effect of TGF- β 1 on myodifferentiation, and most likely accounts for the undifferentiated appearance of the C2(280) grafts.

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